

In-silico prediction of microbial assemblages

from metagenomics to community modelling

Giovanni Bacci - giovanni.bacci@unifi.it





















































































Microbes matter

Microbes are a lot! They are (almost) everywhere They do a lot of things



Host genome integration (Hologenome)



Useful compounds

Biogeochemical cycles





- Who is there?
- How are they influencing each other?



- doing?
- What are they doing?
 - How fast are they growing?

- Who is there?
- How are they influencing each other?



- doing
- What are they doing?
 - How fast are they growing?



- Who is there?
- How are they influencing each other?





- What are they doing?
- How fast are they growing?

• What are they doing? • Who is there? • How fast are they growing? • How are they influencing each other?

How can we model such complex interactions?



S = n° taxa

Community sampling



S = n° taxa

Community sampling



S = n° taxa

Taxonomic profiling

Community sampling



. . .

S = n° taxa

Taxonomic profiling

target gene (taxonomic marker) shotgun metagenomics T-RFLP DGGE

Community sampling



. . .

S = n° taxa

Taxonomic profiling

target gene (taxonomic marker) shotgun metagenomics T-RFLP DGGE



 DNA sequencing
 Identify common genes within a community
 Identify genome contents favored by current environmental conditions

Community sampling



. . .

S = n° taxa

How can we use quantitative information to infer community structure?

Taxonomic profiling

target gene (taxonomic marker) shotgun metagenomics T-RFLP DGGE



DNA sequencing • Identify common genes within a community • Identify genome contents favored by current environmental conditions









- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error
- Nodes are ranked according to their prediction power



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error
- Nodes are ranked according to their prediction power
- The system is optimised using genetic algorithm



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error
- Nodes are ranked according to their prediction power
- The system is optimised using genetic algorithm



- All nodes (taxa) are predicted using all other nodes in the network
- The effect of each predictor is estimated using root mean squared error
- Nodes are ranked according to their prediction power
- The system is optimised using genetic algorithm





Preliminary results

R² value ranging from 0.65 to 1 (except for sp7)

Thaiss et al 2016 - Persistent microbiome alterations modulate the rate of post-dieting weight regain

What about metabolism?











- Nutrient effect simulation
- Effect of antibiotic molecules
- Changes in metabolic assets



- Nutrient effect simulation
- Effect of antibiotic molecules
- Changes in metabolic assets



- Nutrient effect simulation
- Effect of antibiotic molecules
- Changes in metabolic assets





- Nutrient effect simulation
- Effect of antibiotic molecules
- Changes in metabolic assets





- Nutrient effect simulation
- Effect of antibiotic molecules
- Changes in metabolic assets

⇒ Biomass = $f(A) = \mu(h^{-1})$ ⇒ Coverage = n_{reads}

Coverage = *f*(*Biomass*)



Growth rates estimation from metagenomes

Korem et al 2015 - Growth dynamics of gut microbiota in health and disease inferred from single metagenomic samples





During stationary phase coverage is evenly distributed along the entire genome





During exponential phase coverage increases near the origin of replication





During exponential phase coverage increases near the origin of replication



2^{ori} PTR **)**ter



During exponential phase coverage increases near the origin of replication





 $PTR = \frac{2^{ori}}{2^{ter}} = \text{coverage close to the origin of replication}$

During exponential phase coverage increases near the origin of replication



 $PTR = \frac{2^{ori}}{2^{ter}} = \text{coverage close to the origin of replication}$ = coverage close to the termination

Growing bacterial population





 PTR can be inferred from the same data used for community profiling

Growing bacterial population





- PTR can be inferred from the same data used for community profiling
- Reference complete genome must be available

Growing bacterial population





- PTR can be inferred from the same data used for community profiling
- Reference complete genome must be available
- PTR and Growth rate are highly correlated

Growing bacterial population







- PTR can be inferred from the same data used for community profiling
- Reference complete genome
 must be available
- PTR and Growth rate are highly correlated
- We need to find a relation between different growth rate indexes, PTR and coverage

Growing bacterial population









Other preliminary results

In two different conditions (NC and HFD) PTR and growth rate seem to be correlated



Other preliminary results

In two different conditions (NC and HFD) PTR and growth rate seem to be correlated

THANKS FOR THE ATTENTION!

THANKS FOR THE ATTENTION!

Questions, suggestions, and collaborations are welcome